CSU IBC Meeting - Approved Minutes

Meeting title	CSU IBC Meeting September 2026
Date	September 9, 2026
Convened at	8:31am
Location	USC 320
Open or closed meeting (If closed, why?)	The meeting was closed briefly to discuss personal, proprietary, and/or security matters. The meeting was open the remainder of the time.
Review of prior meeting minutes	August 12, 2026 IBC meeting minutes approved as written
Adjournment	9:47am
Recorded by	M. Ramey & C. Johnson

Attendees

Check if Attending (Members):	**Check if Attending (Alternate Members):**
☒ Jessica Ayers, Animal expert	☐ Lon Kendall, Director LAR
☐ Donald Bade, Unaffiliated	
☒ Angela Bosco-Lauth, Chair	
☒ Joshua Chan, Synthetic Biology	
☒ Chaoping Chen, Retrovirology	
☒ Jason Cummings, Lab Representative	
☒ Marc Nishimura, Plant Expert	
☒ Brendan Podell, Mycobacteria specialist	
☐ Ann Powers, Virology	
☒ Robyn Roberts, Assoc Chair/Plant expert	
☒ Joanie Ryan, Assistant Biosafety Director	☒ Rebecca Moritz, Biosafety Office Director*
☐ Carrie Satterwhite, Unaffiliated	☒ Peter O'Brien, Alternate Unaffiliated
☐ Tony Schountz, Virology	☒ Christine Johnson, IBC/IRE Manager, Alternate-at-Large*
Quorum = 7 voting members; _10_ in attendance	*non-voting at this meeting
Non-Voting Members:	
☒ Joni Van Sickle, Occupational Health Coord	
☐	
ORCC Staff (non-voting):	
☐ Michelle Ramey, Assistant Compliance Coord.	☒ Sonia Aleman Rivera, Assistant Biosafety Officer
☒ Nicole Marlenee, Biosecurity Specialist	☒ Kelly Kim, Assistant Biosafety Officer
☒ Scott Van Scotter, Biosafety Trainer	
Additional Guests:	
Mario Arango, OGC	

Quorum was maintained throughout. Any member with a conflict of interest left the meeting during discussion and/or committee determination on the conflicted items.

1. Review of August 12, 2026 IBC meeting minutes

The meeting minutes from August 12, 2026 were reviewed and approved as written.

2. Agent/Project Review

a. PI name/Lab	**Myers Lab (tabled last meeting)**
Project Number and Title	**26-039B: Cell type-specific retrograde tracing in the nervous system**
Project Overview	Glycoprotein-deleted rabies virus (RVΔG) is a specially modified virus used to map how neurons connect in the brain. It's not a live virus that causes rabies; it's an engineered tool for studying brain circuits. For these studies, we will administer a combination of viruses to examine the brain-wide inputs to a population of hypothalamic dopamine neurons.
Planned Modifications	Genetic Modification of RV: -The rabies glycoprotein (G), essential for viral budding and trans-synaptic spread, is deleted from the viral genome. -A transgene (e.g., green fluorescent protein [GFP]) replaces the G gene, enabling visualization of infected cells.
Agents used	• Glycoprotein-deleted rabies virus (RVΔG)
NIH Guidelines Section	III-D-1; III-D-4
Proposed containment conditions (BSL, ABSL, etc.)	BSL2/ABSL2
Discussion	1. The IBC recommens to distinguish between PPE for bench work and for surgery 2. The rat animal model associated to this project is marked as being genetically modified. However, there is no mention of this in the project. Please briefly describe how the genetic modifications contribute to the project.
Verification of training	All training is up to date.
Verification of facilities	The Lab is up to date on inspections.
Motion	A motion was made to approve the project with modifications described above (see Discussion).
Vote	The motion was unanimously approved.
b. PI name/Lab	**Kading Lab**
Project Number and Title	**26-041B: Mechanisms underpinning mosquito vector competence for Rift Valley fever and Japanese encephalitis viruses**
Project Overview	This project will evaluate the involvement of major immune pathways in transmission of both RVFV and JEV across multiple mosquito vectors and tissues, with the goal of characterizing immune modulators or pathways commonly associated with transmission as well as those unique to particular vectors, viruses, or tissues. We will additionally use molecular and microscopy approaches to investigate the expression of potential

	receptors to virus entry into the mosquito ovaries for vertical transmission.
Planned Modifications	dsRNA will be used to knockdown gene expression in mosquitoes
Agents used	• Rift Valley Fever virus (RVFV) • Japanese encephalitis virus
NIH Guidelines Section	III-D-4
Proposed containment conditions (BSL, ABSL, etc.)	BSL3
Discussion	1. Briefly describe how knockdown will be confirmed. 2. Confirm whether the mosquitoes will be undergoing genetic modifications, please update this in the Source Materials section appropriately. 3. Please work with biosafety on updating FSAP work objectives to cover this new project.
Verification of training	All training is up to date.
Verification of facilities	The Lab is up to date on inspections.
Motion	A motion was made to approve the project with modifications described above (see Discussion).
Vote	The motion was unanimously approved.

3. Amendments requiring full IBC review

c. PI name/Lab	**Lark Lab**
Project Number and Title	**26-004B: Tumor-mediated trafficking of extracellular vesicles**
Project Overview	The goal of this study is to define how tumors contribute to cachexia in adipose tissue and skeletal muscle through release and delivery of extracellular vesicles (EVs). Specifically, we will inject murine cancer cell lines into mice then track the extracellular vesicles released.
Planned Modifications	Transduce cell lines and then inject in mice.
Agents used	• Lentivirus particles
NIH Guidelines Section	III-D-3; III-D-4
Current containment conditions (BSL, ABSL, etc.)	BSL2/ABSL2
Amendment request	Request to lower ABSL to ABSL1
Discussion	In the Source Materials section, update the biosafety level for the mouse model to ABSL1.
Verification of training	Training up to date for all Lab members.
Verification of facilities	The Lab is up to date on inspections.

Motion	A motion was made to approve the project with modifications described above (see Discussion).
Vote	The motion was unanimously approved.
d. PI name/Lab	**Kim Lab**
Project Number and Title	**17-030B: Investigation of neuronal activity**
Project Overview	We will use AAV to express neuronal activity indicator or proteins that regulate neuronal activity in cultured neurons or mouse brains to monitor neuronal activity. We will infect neurons with AAVs in vitro or in vivo via stereotaxic injection
Planned Modifications	AAV vectors expressing various proteins injected into cultured neurons and mouse brains
Agents used	• AAV • Lentivirus vector
NIH Guidelines Section	III-D-4
Proposed containment conditions (BSL, ABSL, etc.)	BSL1/ABSL1
Amendment request	Adding AAV vectors
Discussion	1.The Project Summary mentions using Lentivirus, however this is not described in the Experimental and Procedural Details. If the Lentivirus work is not conducted in this project, please remove it from the summary. If it is used, please add details regarding what and how Lentivirus is used, including PPE and containment procedures. 2. In the description of PPE use, the language should be changed to indicate eye protection will be used at all times in the lab. 3. Verify all other PPE. Specifically, are scrubs and surgical masks always worn when working with the vector or only for surgeries? Please clarify this. 4. Under disinfectant, please add actual sodium hypochloride concentration, this should be freshly prepared. 5. Please review and update the sequences/genes table to include newly added sequence/genes. 6. Please update the “Brief Summary of Project” section to include the purpose of the project and the overall methodology approach. 7. Many details are missing from the “Description of Experimental and Procedural Details”. Please describe the methodology for the AAV infection. Which neuronal proteins will be expressed? Will infections be done in vitro and in vivo? What serotypes will be used? 8.It is unclear if live animals are used for infections and imaging. Are stereotaxic injections being done in this project? How will mice be handled and contained? Where will the injection take place? Please add details on how imaging will be done.

	9. Please add PPE used in the project and describe decontamination and disposal procedures. 10. Please associate the appropriate biological materials to the appropriate projects. 11. Please add additional project personnel to project: Using Lentiviral vector to express delta-catenin in neurons. 12. As per the CSU Eye Protect Policy, appropriate eye protection is required at all times in the labs. Please update the language in the project to reflect this requirement.
Verification of training	Training up to date for all Lab members.
Verification of facilities	The Lab is up to date on inspections.
Motion	A motion was made to table the project until additional information/modifications described above (see Discussion) is received. Can re-review via email if there is an urgent need.
Vote	The motion was unanimously approved.
e. PI name/Lab	**Clay Lab**
Project Number and Title	**18-060B: Recombinant AAV delivery to sheep hypothalamus and pituitary**
Project Overview	The hypothesis underlying this proposal is that adeno-associated virus (AAV) will be an effective delivery vector to target transgene expression in specific neuronal populations in the ovine central nervous system and the ovine pituitary. Our specific objective is to demonstrate that in vivo delivery of a recombinant AAV will allow for GnRH neuronal-specific expression of a fluorescent protein and pituitary gonadotrope-specific expression of a fluorescent protein.
Planned Modifications	AAV delivering CRISPRi constructs will be injected into ovine pituitary
Agents used	• AAV
NIH Guidelines Section	III-D-4
Proposed containment conditions (BSL, ABSL, etc.)	BSL1/ABSL1
Amendment request	Adding AAV vectors and intravascular delivery to the pituitary
Discussion	The lab is requesting to add a delivery method and additional AAV vector. No substantial change, the overall scrope remains the same. The appropriate PPE and containment are listed. The IBC had no concerns.
Verification of training	Training up to date for all Lab members.
Verification of facilities	The Lab is up to date on inspections.
Motion	A motion was made to approve the amendment request as written.
Vote	The motion was unanimously approved.

f. PI name/Lab	**Scountz Lab**
Project Number and Title	**25-064B: Experimental infection of deer mice with Andes viruses**
Project Overview	Project amended 9/10/26: We will test a new vaccine platform using H18N11 influenza A virus encoding Andes virus Gn or Gc spike proteins to determine immunogenicity, neutralizing antibody response, and protection from infection upon challenge with Andes virus in deer mice.
Planned Modifications	Testing a recombinant vaccine in mice
New Agents	• Alphavirus replicon (Semliki Forest virus) • H18N11 influenza A vaccine platform, hantavirus glycoproteins
NIH Guidelines Section	III-D-4
Proposed containment conditions (BSL, ABSL, etc.)	BSL2/ABSL2
Amendment request	Adding vaccine agent
Discussion	1. It was assumed that the agent H18N11 influenza A vaccine platform, hantavirus glycoproteins was meant to be included in the project. If this is correct, please add. 2. Please provide more details regarding the vaccine platforms. How is vaccine platforms created? Will they be made in the lab or acquired from an outside source? 3. Please specify the PPE used for the BSL2 portion of this work.
Verification of training	Training up to date for all Lab members.
Verification of facilities	The Lab is up to date on inspections.
Motion	A motion was made to approve the project with modifications described above (see Discussion).
Vote	The motion was unanimously approved.

4. New Business

a. **Draft NIH Biosafety Policy for Research Involving Biohazards – for open comment** (https://grants.nih.gov/grants/guide/notice-files/NOT-OD-26-112.html)
The IBC/IRE Manager gave an overview of the NIH summary table with the proposed changes and their potential impact on CSU research and the IBC. There were several concerns regarding the broadening scope, as well as privacy and security concerns related to the proposed incident reporting requirements. CSU is working on an institutional response, and IBC members were also encouraged to submit comments of their own. The NIH comment period ends on October 19th.

b. **Review of Updated Biosafety Manual**
The Biosafety Office is working on updating the Biosafety Manual to bring it up to date and improve readability. The committee was asked to review the document prior to the next meeting, so that the committee could vote on it then.

5. Unfinished Business

a. U.S. Government Policy for Stopping High-Risk Life Sciences Research
The IBC/IRE Manager and IRE Chair have been meeting with folks in Sponsored Programs and with OVPR leadership to determine what this process will look like. Additionally, the updated DGOF policy is being drafted.

6. Closed Session
Items to discuss

7. Reports

a. Coordinator's Report	1. Next IBC meeting: Wednesday, October 14, 2026 2. IBC/IRE Coordinator position has been posted. This person will report to the IBC/IRE Manager and will assist with IBC and IRE functions. Please help circulate the job posting.
b. Biosafety Office Report	1. Inspections update: no update 2. Incident Reports: no incidents involving rDNA/synthetic nucleic acids were reported.

8. Public Comment
None

9. Items to be read into the minutes

a. Items Reviewed at Previous IBC Meeting and Approved After Modification were completed	a. **McGrath Lab** **Project: 26-033B** - Assessing Safety of Bioengineered EcN 1917 as a Bacterial Living Logic Cancer Therapy; BSL1/ABSL-1. NIH Guidelines category non-exempt rDNA: III-D-4 b. **Woerman Lab** **Project: 23-039B -** Investigating prion strain biology; BSL2+/ABSL-2+. NIH Guidelines category non-exempt rDNA: III-D-2, III-D-4 c. **Peebles Lab** **Project:** 15-092B - Expression of proteoglycans in Wolffia australiana and Wolffia globosa (duckweeds). BSL1. NIH Guidelines category non-exempt rDNA: III-E-2